

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
4 April 2002 (04.04.2002)

PCT

(10) International Publication Number
WO 02/27316 A2

(51) International Patent Classification⁷: **G01N 33/53**

LACH, Agnieszka; 915 North Racine, Unit 1SW, Chicago, IL 60022 (US). **PESTEL, Cynthia, D.**; 131 South Luther Avenue, Lombard, IL 60148 (US). **RAMP, John, M.**; 462 Pine Grove Street, Gurnee, IL 60031 (US).

(21) International Application Number: **PCT/US01/29390**

(22) International Filing Date:

20 September 2001 (20.09.2001)

(74) Agents: **CASUTO, Dianne** et al.; Abbott Laboratories, 100 Abbott Park Road, D377 AP6D/2, Abbott Park, IL 60064-6050 (US).

(25) Filing Language:

English

(26) Publication Language:

English

(81) Designated States (*national*): CA, JP.

(30) Priority Data:

09/669,082 25 September 2000 (25.09.2000) US

(84) Designated States (*regional*): European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR).

(71) Applicant: **ABBOTT LABORATORIES** [US/US]; D377 AP6D, 100 Abbott Park Road, Abbott Park, IL 60064-6050 (US).

Published:

— without international search report and to be republished upon receipt of that report

(72) Inventors: **SCOPP, Richard, L.**; 4803-68th Street, Kenosha, WI 53142 (US). **FINLEY, David, M.**; 2414-S. Hidden Trail, Spring Grove, IL 60081 (US). **TRIMPE, Kevin, L.**; 501 Majestic Court, Gurnee, IL 60031 (US).

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 02/27316 A2

(54) Title: METHODS AND KITS FOR DECREASING INTERFERENCES IN PLASMA OR SERUM CONTAINING ASSAY SAMPLES OF SPECIFIC BINDING ASSAYS

(57) Abstract: Methods and kits are provided for decreasing interferences and inaccuracies due to nonoptimal sample handling of blood samples in plasma or serum containing assay samples of specific binding assays by addition of a large polycation to the assay sample during the specific binding assay.

BEST AVAILABLE COPY

METHODS AND KITS FOR DECREASING INTERFERENCES IN PLASMA OR SERUM CONTAINING ASSAY SAMPLES OF SPECIFIC BINDING ASSAYS

Field of the Invention

The present invention relates to an improved method for performing specific binding assays with plasma or serum samples wherein a relatively large polycation is added to the assay sample during the assay. The present invention also relates to improved specific binding assay kits for plasma or serum samples which comprise as one component of the kit a solution containing a large polycation.

Background of the Invention

Polycations are organic or inorganic, synthetic or naturally occurring, compounds having at least two positive charges. Examples of relatively large polycations include, but are not limited to, polylysine, polyethyleneimine and polypropyleneimine and their lower alkyl ammonium salts such as polybrene, and MERQUAT.

Polycations such as polylysine, polyarginine and polyhistidine are commercially available for use as enzyme inhibitors, as substrates in the isolation of plasma membranes, in chromosomal preparations, in microencapsulation, in sustained release delivery devices, and as drug delivery devices. Poly-L-lysine is also used as a carrier protein in the synthesis of immunogens, while poly-D-lysine is used as a carrier protein in immobilized antigen enzyme linked immunosorbent assays (ELISAs). Polycations such as poly(N-ethyl-4-vinylpyridinium have also been used, in conjunction with polyanions such as poly(methacrylate), as carriers for reactants in both ELISAs (Yazynina et al. Analytical Chemistry 1999 71(16):3538-43) and visual enzyme immunoassays (Dzantiev et al. Immunology Letters 1994 41(2-3):205-11).

Polyionic reagents including polycations have been disclosed for use in initiating non-specific binding of a substance to magnetic particles. For example, U.S. Patents 4,935,147, 5,076,950, 5,279,936 and 5,770,388 disclose a list of exemplary polycationic reagents including polyalkylene amines such as polyethyleneimine and polypropyleneimine

and their lower alkyl ammonium salts such as polybrene $(N(CH_3)_2CH_2CH_2N(CH_3)_2CH_2CH_2CH_2CH_2)_n$, metal ions such as calcium and barium ions, aminodextrans, protamine, positively charged liposomes, polylysine, and the like for use as a chemical means for forming non-specific bonds between the substance and magnetic particles.

Polycations have also been taught to be useful in separation techniques for immunoassay of whole blood samples. WO 9936781 discloses a chromatography assay device which separates red blood cells in a sample from serum or plasma prior to movement of the serum or plasma down the chromatography column. The red blood cell separating agent used in this device is preferably a polycation comprising poly-L-lysine hydrobromide, poly-L-arginine hydrochloride, poly-L-histidine, poly(lysine, alanine) 3:1 hydrobromide, poly(lysine, arginine) 2:1 hydrobromide, poly(lysine, alanine) 1:1 hydrobromide, poly(lysine, tryptophan) 1:4 hydrobromide or particularly poly(diallyldimethylammonium chloride). However, addition of a separating agent such as a polycation directly to the assay system is taught to interfere with the system, often by aggregating other reagents and binding members in addition to the red blood cells.

Accordingly, an object of the present invention is to provide a method for decreasing interferences which result in inaccurate readings in plasma or serum containing assay samples of specific binding assays. The method comprises adding a large polycation to the plasma or serum containing assay sample during the specific binding assay.

Another object of the present invention is to provide improved specific binding assay kits for plasma and serum containing assay samples which comprise as one component of the kit a solution containing a large polycation.

Summary of the Invention

The present invention provides a method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of specific binding assays comprising adding an effective amount of a large polycation to serum or plasma containing assay samples during the specific binding assay. In a preferred embodiment, the large polycation has a molecular weight of 3,000 daltons or greater. In another preferred embodiment, the large polycation is a polylysine, polyornithine, polybrene or MERQUAT.

In a more preferred embodiment, the large polycation comprises a polylysine with a molecular weight ranging between 5,200 and 11,200 daltons. In another more preferred embodiment, the large polycation comprises polylysine with a molecular weight of 8,800 daltons. In another preferred embodiment, the specific binding assay is performed on a solid phase, such as paramagnetic microparticles. In other embodiments, the specific binding assay measures thyroid stimulating hormone, free prostate specific antigen (PSA), alpha fetal protein, hepatitis B core antibody, hepatitis B surface antibody or human immunodeficiency virus.

The invention also provides a method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a thyroid stimulating hormone specific binding assay comprising adding a large polycation to serum or plasma containing assay samples during the thyroid stimulating hormone specific binding assay. In a preferred embodiment, the large polycation has a molecular weight of 3,000 daltons or greater. In another preferred embodiment, the large polycation is a polylysine, polyornithine, polybrene or MERQUAT. In a more preferred embodiment, the large polycation comprises a polylysine with a molecular weight ranging between 5,200 and 11,200 daltons. In another more preferred embodiment, the large polycation comprises polylysine with a molecular weight of 8,800 daltons. In another preferred embodiment, the specific binding assay is performed on a solid phase, such as paramagnetic microparticles. In a most preferred embodiment, the thyroid stimulating hormone specific binding assay comprises:

- a) forming a first complex by incubating a serum or plasma sample with paramagnetic microparticles coated with anti- β TSH antibody and an assay diluent which comprises a large polycation, for a time and under conditions which allow the thyroid stimulating hormone present in the sample to bind to the anti- β TSH antibody coated microparticles;
- (b) forming a second complex by incubating the first complex with an acridinium labeled conjugate comprising an anti- α TSH antibody, for a time and under conditions which allow the conjugate to bind to the first complex;
- (c) creating a chemiluminescent reaction in the second complex; and

(d) measuring the chemiluminescent reaction as relative light units wherein the amount of thyroid stimulating hormone in the plasma or serum sample is directly related to the measured relative light units.

The present invention also provides a method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a free or total prostate specific antigen specific binding assay comprising adding a large polycation to serum or plasma containing assay samples during the free or total prostate specific antigen specific binding assay. In a preferred embodiment, the large polycation is a polylysine or polyornithine. In another preferred embodiment, the free prostate specific antigen (PSA) specific binding assay comprises:

(a) forming a first complex by incubating a serum or plasma sample with paramagnetic microparticles coated with an antibody specific for free PSA, for a time and under conditions which allow the free PSA present in the sample to bind to the antibody coated microparticles;

(b) forming a second complex by incubating the first complex with an acridinium labeled conjugate comprising an anti-PSA antibody, for a time and under conditions which allow the conjugate to bind to the first complex;

(c) creating a chemiluminescent reaction in the second complex; and

(d) measuring the chemiluminescent reaction as relative light units wherein the amount of prostate specific antigen in the plasma or serum sample is directly related to the measured relative light units.

In another preferred embodiment the total PSA specific binding assay comprises:

(a) forming a first complex by incubating a serum or plasma sample with paramagnetic microparticles coated with an antibody which binds both free and complexed PSA, for a time and under conditions which allow the PSA present in the sample to bind to the antibody coated microparticles;

(b) forming a second complex by incubating the first complex with an acridinium labeled conjugate comprising an anti-PSA antibody, for a time and under conditions which allow the conjugate to bind to the first complex;

(c) creating a chemiluminescent reaction in the second complex; and

(d) measuring the chemiluminescent reaction as relative light units wherein the amount of prostate specific antigen in the plasma or serum sample is directly related to the measured relative light units.

The present invention also provides an improved specific binding assay kit for plasma and serum samples comprising a solution containing a large polycation. In a preferred embodiment, the large polycation has a molecular weight of 3,000 daltons or greater. In another preferred embodiment, the large polycation is a polylysine, polyornithine, polybrene or MERQUAT. In a more preferred embodiment, the improved specific binding assay kit comprises a specific binding assay which measures thyroid stimulating hormone, free prostate specific antigen, alpha fetal protein, Hepatitis B core antibody, Hepatitis B surface antibody or human immunodeficiency virus.

The present invention also provides an improved kit for detection of thyroid stimulating hormone comprising:

- (a) mouse, monoclonal anti- β TSH coated microparticles;
- (b) mouse, monoclonal anti- α TSH acridinium-labeled conjugate; and
- (c) a modified TSH assay diluent comprising a large polycation. Preferably, the large polycation is a polylysine having a molecular weight from 5,200 to 11,200 daltons.

The present invention also provides an improved kit for detection of free prostate specific antigen comprising:

- (a) microparticles comprising a monoclonal antibody specific to free PSA in a diluent comprising a large polycation;
- (b) mouse, monoclonal anti-PSA acridinium-labeled conjugate. Preferably, the large polycation is a polylysine or polyornithine.

Detailed Description of the Invention

Non-optimal serum or plasma sample preparation techniques including, but not limited to, inadequate centrifugation, incomplete clotting time, and exposure to thermal stress, have been found to cause interferences in plasma or serum containing assay samples which lead to inaccurate readings in specific binding assays. It has now been found that addition of a large polycation to a plasma or serum containing assay sample during the

specific binding assay decreases or eliminates these interferences so that accurate readings can be obtained.

For purposes of the present invention, by "large" polycation it is meant a polycation with a molecular weight of approximately 3,000 daltons or greater. Examples of large polycations useful in the present invention include, but are not limited to, polylysines with a molecular weight ranging between 5,200 and 11,200, polyornithine with a molecular weight of 5300, polybrene with a molecular weight ranging between approximately 4,000 and 6,000 daltons, and MERQUAT with a molecular weight of approximately 4,000,000 daltons. The polycation can be added during the immunoassay as a separate reagent. Alternatively, the polycation can be incorporated into an assay specific diluent.

The amount of polycation used in an assay may vary depending on the type and its molecular weight. Generally, however, the amount used is a quantity which is effective at achieving the desired result, i.e. eliminating interference, without detrimentally affecting other assay parameters (such as sensitivity, specificity, etc.). By way of example, polycations such as polylysines, polyornithines, polyarginines, and polyhistidines at final concentrations ranging from about 0.005% to about 1% weight/volume (wt/vol) may be used. More preferably, polylysines ranging from about 0.01% to about 0.5% wt/vol are used. Even more preferably, polylysines ranging from about 0.1% to about 0.5% wt/vol are used. For a polylysine with a molecular weight of 8,800 daltons, a concentration of about 0.25% is preferred. For polybrene, concentrations ranging from about 0.2% to 1% wt/vol are preferred. For MERQUAT, concentrations ranging from about 0.15% to about 0.30% are preferred.

While higher concentrations of a polycation may still be effective at decreasing interferences in the sample, it is believed that the higher viscosity resulting from addition of some polycations may cause carryover, particularly in high throughput automated specific binding assay systems. However, those of ordinary skill in the art could easily determine the proper concentration suitable for a particular assay.

The polycations of the present invention may be used in any type of specific binding assay that tests for the presence of an analyte (such as an antigen or antibody) in a serum or plasma sample, including but not limited to sandwich and competitive type

immunoassays. Such immunoassays may utilize reagents comprising a polyclonal or monoclonal antibody, fragments of said antibodies (such as an Fab'2 fragment) or combinations of polyclonal, monoclonal and antibody fragments. Typically in such assays, a labeled reagent (such as a labeled antigen or antibody) is used for detecting and/or quantitating an analyte of interest. Such labels include, without limitation, enzymatic, fluorescent, chemiluminescent, and radioactive labels. The manner of making and using all types of immunoassays as well as the reagents and/or labeled reagents used in such assays are well known to routine practitioners in the art.

One embodiment of the present invention relates to an improved specific binding assay for measuring TSH in serum or plasma samples. In a preferred embodiment, the TSH specific binding assay comprises a modified ARCHITECT TSH assay format (Abbott Laboratories, Abbott Park, IL 60035-6050) wherein a large polycation with a molecular weight of approximately 3,000 daltons or greater is added to the assay sample during the assay, i.e. before or during the incubation of the sample with the solid phase. In this embodiment, it is preferred that the polycation be a polylysine with a molecular weight ranging between 5,200 and 11,200 daltons, with a polylysine having a molecular weight of approximately 8,800 daltons being preferred. It is also preferred that the polycation be incorporated within the TSH assay diluent which is combined with the plasma or serum sample and the TSH antibody. Preferred concentration ranges of polylysine in the TSH assay range from about 0.1% to about 1% wt/vol with 0.25% wt/vol being most preferred.

Another embodiment of the present invention relates to improved kits for performing this modified ARCHITECT TSH assay. Kits of the present invention comprise at least anti- β TSH (mouse, monoclonal) coated microparticles in a buffer, preferably TRIS buffer, and even more preferably with protein (bovine) stabilizers and antimicrobial agents as a preservative, an acridinium-labeled conjugate comprising a mouse anti- α TSH monoclonal antibody, preferably in MES (2-[N-Morpholino]ethanesulfonic acid) buffer with protein (bovine) stabilizers and antimicrobial agents as a preservative; and a modified TSH assay diluent comprising a buffer, preferably TRIS, containing a polycation, preferably a polylysine ranging in molecular weight from 5,200 to 11,200 daltons at a concentration ranging from about 0.1% wt/vol to about 0.5% wt/vol. It is preferred that this diluent comprise antimicrobial agents as preservatives. Alternatively, the polycation can be

provided as a separate kit component for addition to the assay samples along with the TSH assay diluent. Kits of this embodiment of the present invention may also comprise a Multi-Assay Manual Diluent containing phosphate buffered saline solution with an antimicrobial agent as a preservative; a Pre-Trigger Solution containing 1.32% (w/v) hydrogen peroxide; a Trigger Solution containing 0.35 N sodium hydroxide; and a wash buffer containing phosphate buffered saline solution and an antimicrobial agent preservative.

A second preferred embodiment of the present invention relates to an improved specific binding assay for measuring free or total prostate specific antigen (PSA) in serum or plasma samples. In a most preferred embodiment, the PSA specific binding assay comprises a modified ARCHITECT total or free PSA assay format (Abbott Laboratories, Abbott Park, IL 60035-6050) wherein a large polycation with a molecular weight of approximately 3,000 daltons or greater is added to the assay with the assay sample, i.e. before or during the incubation of the sample with the solid phase.

In this embodiment, it is preferred that the polycation be a polylysine with a molecular weight ranging between 5,200 and 11,200 daltons. It is also preferred that the polycation be incorporated in the diluent of the anti-PSA coated microparticles (hereinafter "microparticle diluent") which is combined with the plasma or serum sample. Preferred concentration ranges of polylysine in the total PSA assay range from about 0.005% to about 1% wt/vol with 0.005% wt/vol being most preferred. Preferred concentration ranges of polylysine in the free PSA assay range from about 0.01% to about 1% wt/vol with 0.01% wt/vol being most preferred.

Another embodiment of the present invention relates to improved kits for performing a modified ARCHITECT total or free prostate specific antigen (PSA) assay. A kit of the present invention comprises microparticles, coated with an anti-PSA monoclonal antibody (one that is specific for free PSA in the case of the free PSA assay and one that binds both free and complexed PSA for the total PSA assay) in a diluent which also contains a polycation. The kit also includes an acridinium-labeled conjugate comprising an anti-PSA monoclonal antibody. Preferably, the polycation is a polylysine ranging in molecular weight from about 5,200 to about 11,200 daltons at a concentration ranging from about 0.005% wt/vol-0.5% wt/vol. The buffer of the microparticle diluent preferably is a TRIS buffer and even more preferably contains protein (bovine) stabilizers and

antimicrobial agents as a preservative. The acridinium-labeled conjugate is preferably in MES (2-[N-Morpholino]ethanesulfonic acid) buffer with protein (bovine) stabilizers and antimicrobial agents as a preservative. Alternatively, the polycation can be provided as a separate kit component for addition to the assay samples along with the PSA microparticle diluent. Kits of this embodiment of the present invention may also comprise a Pre-Trigger Solution containing 1.32% (w/v) hydrogen peroxide, a Trigger Solution containing 0.35 N sodium hydroxide, and a wash buffer containing phosphate buffered saline solution and an antimicrobial agent preservative.

Re-centrifugation of nonoptimally handled plasma and serum samples has also been demonstrated to be effective in decreasing interferences and restoring sensitivity and accuracy in sample measurement in specific binding assays for alpha fetal protein (AFP), Hepatitis B core antibody (HBcAb), Hepatitis B surface antibody (HBsAb), and human immunodeficiency virus (HIV). Accordingly, it is believed that addition of a large polycation to plasma or serum containing assay samples during performance of specific binding assays for these analytes will also be useful in decreasing interferences due to nonoptimal sample preparation.

The following nonlimiting examples are provided to further illustrate the present invention.

EXAMPLES

Example 1: Preparation of Contaminated Plasma or Serum Samples

Blood was drawn from one volunteer into four serum separator tubes, also referred to as SST Vacutainer tubes (Becton Dickinson, Number 366510) and six ethylenediaminetetracetic acid (EDTA) Vacutainer tubes (Becton Dickinson, Number 366457). The blood was allowed to clot for 30 minutes and then spun in a centrifuge at 3,500 RPM for 10 minutes. Serum was recovered from the four SST tubes. Plasma was recovered from the six EDTA tubes. A portion of the plasma was then contaminated by addition of 60 microliters of buffy coat (including red blood cells) from the EDTA tubes.

Example 2: Effect of Polycations in the ARCHITECT TSH Assay

a. General Procedure: The ARCHITECT TSH assay (Abbott Laboratories, Abbott Park, IL. 60035-6050) is a two-step immunoassay which determines the presence of thyroid stimulating hormone (TSH) in human serum and plasma using Chemiluminescent Microparticle Immunoassay (CMIA) technology with flexible assay protocols, referred to as CHEMIFLEX. In the first step, a serum or plasma sample, anti- β TSH antibody coated paramagnetic microparticles, and TSH Assay Diluent are combined. (The TSH Assay diluent contains 0.5 M TRIS HCl, 1.5 M TRIS base, 1.3 M NaCl, 0.2% of the antimicrobial agent NIPASEPT (Nipa Laboratories Ltd., Wilmington DE) and the antimicrobial agent A56620 (Abbott Laboratories, Abbott Park, IL), at pH 8.8). TSH present in the sample binds to the anti-TSH antibody coated microparticles. After washing, anti- α TSH acridinium labeled conjugate is added as the second step. Two solutions referred to as a Pre-Trigger and Trigger Solution, which comprise hydrogen peroxide and sodium hydroxide, respectively, are then added to the reaction mixture and the resulting chemiluminescent reaction is measured as relative light units (RLUs). A direct relationship exists between the amount of TSH in the plasma or serum sample and RLUs detected by the ARCHITECT/optical system.

b. Experimental Design: Experiments were designed in which serum and plasma samples were contaminated intentionally with red blood cells to interfere with the sensitivity of the assay (see Example 1). In separate experiments, a polycation, i.e. polylysine, polybrene or MERQUAT, then was added to the TSH Assay Diluent and combined with the serum or plasma sample (150 μ L) and anti- β TSH antibody coated paramagnetic microparticles (50 μ L at 0.1% solids) in the first step of the TSH assay. The assay then was completed as described in the general procedure above.

c. Results: As Table 1 shows polylysines having an average molecular weight of 5,200, 8,800 and 11,200 were found to be effective at eliminating interferences in contaminated samples at a concentration of 0.25%.

Table 1

Type of Polylysine	TSH (uIU/mL) of centrifuged sample	TSH (uIU/mL) of uncentrifuged sample	%Difference
No Polylysine	1.6654	0.1679	90

5200 MW	1.9545	1.9441	1
8800 MW	1.9665	1.9564	1
11,200 MW	1.9939	1.9132	4

Various concentrations of polybrene with a molecular weight of 4,000 to 6,000 daltons also were examined. Concentrations ranging from 0.2% to 1% wt/vol of polybrene were found to be effective at restoring assay sensitivity to contaminated plasma or serum samples without interfering with or altering the functional sensitivity of the TSH assay.

The polycation MERQUAT-100 having a molecular weight of about 4,000,000 daltons also restored assay sensitivity to contaminated samples without interfering with overall function of the assay at concentrations of either 0.15% or 0.30% in the TSH Assay Diluent.

Example 3: Effect of Polycations in the ARCHITECT free PSA Assay

Addition of a polycation to an assay sample also was demonstrated to be effective in decreasing interferences resulting from nonoptimal plasma or serum sample handling in an ARCHITECT free prostate specific antigen (PSA) assay.

a. General Procedure: The ARCHITECT Free PSA assay is a two step immunoassay to determine the presence of free PSA in human serum, using Chemiluminescent Microparticle immunoassay (CMIA) technology. In the first step, a test sample and paramagnetic microparticles, coated with a monoclonal antibody specific to free PSA, are combined. Free PSA present in the sample binds to the anti-free PSA coated microparticles. After washing, anti-PSA acridinium-labeled conjugate is added in the second step. Pre-Trigger and Trigger Solutions are then added to the reaction mixture; the resulting chemiluminescent reaction is measured as RLUs. A direct relationship exists between the amount of free PSA in the sample and the RLUs detected by the ARCHITECT/optical system. Like the TSH assay, nonoptimal preparation of the serum sample leads to interferences in measurement of fluorescence and ultimately an inaccurate reading of the levels of free PSA in the sample.

b. **Experimental Design:** In these experiments, a polycation, in particular, a poly-amino acid, was substituted in place of dextran sulfate in the microparticle diluent. The assay then was performed as described in the general procedure above.

c. **Results:** As shown in Table 2, both polylysine (ranging from 5,200 to 11,200 daltons) and polyornithine (5,300 daltons) at concentrations of 0.025% were effective at decreasing interferences in free PSA measurements caused by poor sample preparation without interfering with or altering the high functional sensitivity of the free PSA assay.

Table 2

Free PSA Concentration (ng/mL)

Sample No.	Dextran Sulfate	poly-L-lysine	poly-L-ornithine	poly-L-arginine	poly-L-histidine
30 (Spun)*	0.699	0.703	0.697	0.528	0.558
30 (Unspun)	0.000	0.666	0.632	0.528	0.511
% Interference**	100%	5%	9%	0%	8%
31 (Spun)	0.617	0.661	0.660	0.462	0.466
31 (Unspun)	0.000	0.614	0.585	0.459	0.441
% Interference	100%	7%	11%	0%	5%

*The term "unspun" refers to an improperly prepared serum or plasma sample which was tested directly in the free PSA assay described above. The term "spun" refers to the same sample, which was re-centrifuged prior to testing.

**% Interference = (Free PSA concentration from Spun sample - Free PSA concentration from Unspun Sample) / (Free PSA concentration from Spun sample) x 100

Although the addition of either polyhistidine (M.W. 13,200 daltons) or polyarginine (M.W. 8,500 daltons) at a concentration of 0.025% also reduced interference, these concentrations interfered with the assay sensitivity. Lower concentrations of these poly-amino acids, however, may be effective at eliminating interference without affecting assay sensitivity.

Example 4: Effect of Polycations in the ARCHITECT total PSA Assay

The general procedure of the ARCHITECT total PSA assay is essentially as described for the free PSA assay in Example 3, with the exception that the paramagnetic microparticles are coated with a monoclonal antibody that binds to both free and

complexed PSA. Experiments were performed in which unspun samples were subjected to a total PSA assay that utilized a microparticle diluent containing dextran sulfate (at a concentration of 0.05%) or a poly-L-lysine of average molecular weight 5200 or 11,200 (in place of dextran sulfate) at a concentration of 0.005%. The results, shown in Table 3, demonstrate that poly-L-lysines of different average molecular weights are effective at decreasing interferences in total PSA measurements in unspun samples without interfering with or altering the high functional sensitivity of the total PSA assay.

Table 3: Unspun Values for Total PSA (ng/mL)

Sample No.	Dextran Sulfate	% Int.	poly-L-lysine 5200 MW	% Int.	poly-L-lysine 11200 MW	% Int.	No poly-L-lysine	% Int.
55	4.070	-62	10.707	-6	10.806	-8	7.724	-27
56	6.663	-33	10.329	-1	10.357	-2	8.922	-8
71	0.083	-99	13.431	-4	14.695	0	1.398	-90
72	0.483	-97	14.797	-8	15.025	-10	5.996	-61
73	0.057	-99	9.983	-8	10.353	-8	1.149	-89
74	0.099	-99	12.483	-1	13.414	4	2.573	-79
75	0.389	-97	14.658	-7	15.211	-6	8.892	-40
76	5.681	-52	12.158	-7	12.158	-5	9.665	-17
Avg. Int.		-80		-5		-4		-51

What is Claimed is:

1. A method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of specific binding assays comprising adding an effective amount of a large polycation to serum or plasma containing assay samples during the specific binding assay.
2. The method of claim 1 wherein the large polycation has a molecular weight of 3,000 daltons or greater.
3. The method of claim 1 wherein the large polycation is a polylysine, polyornithine, polybrene or MERQUAT.
4. The method of claim 3 wherein the large polycation comprises a polylysine with a molecular weight ranging between 5,200 and 11,200 daltons.
5. The method of claim 4 wherein the large polycation comprises polylysine with a molecular weight of 8,800 daltons.
6. The method of claim 1 wherein the specific binding assay measures thyroid stimulating hormone, free prostate specific antigen, alpha fetal protein,, Hepatitis B core antibody, Hepatitis B surface antibody or human immunodeficiency virus.
7. The method of claim 1 wherein said specific binding assay is performed on a solid phase.
8. The method of claim 7 wherein said solid phase comprises paramagnetic microparticles.
9. A method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a thyroid stimulating hormone specific binding assay comprising adding a large polycation to serum or plasma containing assay samples during the thyroid stimulating hormone specific binding assay.

10. The method of claim 9 wherein the large polycation has a molecular weight of 3,000 daltons or greater.
11. The method of claim 9 wherein the large polycation is a polylysine, polybrene or MERQUAT.
12. The method of claim 11 wherein the large polycation comprises a polylysine with a molecular weight ranging between 5,200 and 11,200 daltons.
13. The method of claim 12 wherein the large polycation comprises polylysine with a molecular weight of 8,800 daltons.
14. A method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a thyroid stimulating hormone specific binding assay comprising:
 - a) forming a first complex by incubating a serum or plasma sample with paramagnetic microparticles coated with anti- β TSH antibody and an assay diluent which comprises a large polycation, for a time and under conditions which allow the thyroid stimulating hormone present in the sample to bind to the anti- β TSH antibody coated microparticles;
 - (b) forming a second complex by incubating the first complex with an acridinium labeled conjugate comprising an anti- α TSH antibody, for a time and under conditions which allow the conjugate to bind to the first complex;
 - (c) creating a chemiluminescent reaction in the second complex; and
 - (d) measuring the chemiluminescent reaction as relative light units wherein the amount of thyroid stimulating hormone in the plasma or serum sample is directly related to the measured relative light units.
15. A method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a free prostate specific antigen specific binding

assay comprising adding a large polycation to serum or plasma containing assay samples during the free prostate specific antigen specific binding assay.

16. The method of claim 15 wherein the large polycation is a polylysine or polyornithine.

17. A method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a free prostate specific antigen specific binding assay comprising:

(a) forming a first complex by incubating a serum or plasma sample with paramagnetic microparticles coated with an antibody specific for free PSA, for a time and under conditions which allow the free PSA present in the sample to bind to the antibody coated microparticles;

(b) forming a second complex by incubating the first complex with an acridinium labeled conjugate comprising an anti-PSA antibody, for a time and under conditions which allow the conjugate to bind to the first complex;

(c) creating a chemiluminescent reaction in the second complex; and

(d) measuring the chemiluminescent reaction as relative light units wherein the amount of prostate specific antigen in the plasma or serum sample is directly related to the measured relative light units.

18. An improved specific binding assay kit for plasma and serum samples comprising a solution containing a large polycation.

19. The improved specific binding assay kit of claim 18 wherein the large polycation has a molecular weight of 3,000 daltons or greater.

20. The improved specific binding assay kit of claim 15 wherein the large polycation is a polylysine, polybrene or MERQUAT.

21. The improved specific binding assay kit of claim 18 wherein the specific binding assay measures thyroid stimulating hormone, free prostate specific antigen, alpha fetal

protein, Hepatitis B core antibody, Hepatitis B surface antibody or human immunodeficiency virus.

22. An improved kit for detection of thyroid stimulating hormone comprising:
 - (a) mouse, monoclonal anti- β TSH coated microparticles;
 - (b) mouse, monoclonal anti- α TSH acridinium-labeled conjugate; and
 - (c) a modified TSH assay diluent comprising a large polycation.
23. The kit of claim 19 wherein the large polycation is a polylysine having a molecular weight from 5,200 to 11,200 daltons.
24. An improved kit for detection of free prostate specific antigen comprising:
 - (a) mouse, monoclonal anti-Free PSA coated microparticles in a diluent comprising a large polycation;
 - (b) (b) mouse, monoclonal anti- PSA acridinium-labeled conjugate;
25. The kit of claim 24 wherein the large polycation is a polylysine or polyornithine.
26. A method for decreasing interferences which result in inaccurate readings in serum or plasma containing assay samples of a total prostate specific antigen specific binding assay comprising:
 - (a) forming a first complex by incubating a serum or plasma sample with paramagnetic microparticles coated with an antibody which binds both free and complexed PSA, for a time and under conditions which allow the PSA present in the sample to bind to the antibody coated microparticles;
 - (b) forming a second complex by incubating the first complex with an acridinium labeled conjugate comprising an anti-PSA antibody, for a time and under conditions which allow the conjugate to bind to the first complex;
 - (c) creating a chemiluminescent reaction in the second complex; and

(d) measuring the chemiluminescent reaction as relative light units wherein the amount of prostate specific antigen in the plasma or serum sample is directly related to the measured relative light units.

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
4 April 2002 (04.04.2002)

PCT

(10) International Publication Number
WO 02/27316 A3

(51) International Patent Classification⁷: **G01N 33/53**,
33/543, 33/76, 33/574

South Luther Avenue, Lombard, IL 60148 (US). **RAMP, John, M.**; 462 Pine Grove Street, Gurnee, IL 60031 (US).

(21) International Application Number: PCT/US01/29390

(74) Agents: **CASUTO, Dianne et al.**; Abbott Laboratories, 100 Abbott Park Road, D377 AP6D/2, Abbott Park, IL 60064-6050 (US).

(22) International Filing Date:
20 September 2001 (20.09.2001)

(81) Designated States (*national*): CA, JP.

(25) Filing Language: English

(84) Designated States (*regional*): European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR).

(26) Publication Language: English

(30) Priority Data:
09/669.082 25 September 2000 (25.09.2000) US

Published:

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

(71) Applicant: **ABBOTT LABORATORIES** [US/US];
D377 AP6D, 100 Abbott Park Road, Abbott Park, IL 60064-6050 (US).

(72) Inventors: **SCOPP, Richard, L.**; 4803-68th Street, Kenosha, WI 53142 (US). **FINLEY, David, M.**; 2414-S, Hidden Trail, Spring Grove, IL 60081 (US). **TRIMPE, Kevin, L.**; 501 Majestic Court, Gurnee, IL 60031 (US). **LACH, Agnieszka**; 915 North Racine, Unit 1SW, Chicago, IL 60022 (US). **PESTEL, Cynthia, D.**; 131

(88) Date of publication of the international search report:
20 June 2002

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: METHODS AND KITS FOR DECREASING INTERFERENCES OF ASSAYS SAMPLES CONTAINING PLASMA OR SERUM IN SPECIFIC BINDING ASSAYS BY USING A LARGE POLYCATION

(57) Abstract: Methods and kits are provided for decreasing interferences and inaccuracies due to nonoptimal sample handling of blood samples in plasma or serum containing assay samples of specific binding assays by addition of a large polycation to the assay sample during the specific binding assay.

WO 02/27316 A3

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 01/29390

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 G01N33/53 G01N33/543 G01N33/76 G01N33/574

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

BIOSIS, EP0-Internal, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	US 5 370 993 A (LIN CHENG-I ET AL) 6 December 1994 (1994-12-06) column 17, line 55 -column 18, line 35 examples 4-10,12-16 claims 1-17	1-3,7,8, 18-20 6,9-11, 14-16, 21,22, 24,25
X Y	EP 0 895 085 A (BAYER AG) 3 February 1999 (1999-02-03) the whole document	17,26 15,16, 21,24,25

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *G* document member of the same patent family

Date of the actual completion of the international search

21 March 2002

Date of mailing of the international search report

09/04/2002

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Stricker, J-E

INTERNATIONAL SEARCH REPORT

 Inter national Application No
 PCT/US 01/29390

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 478 729 A (ULLMAN EDWIN F ET AL) 26 December 1995 (1995-12-26) abstract column 6, line 43 - line 53 ---	1
X	DATABASE BIOSIS 'Online! BIOSCIENCES INFORMATION SERVICE, PHILADELPHIA, PA, US; 1995 GOROVITS ELENA L ET AL: "Effect of synthetic polyelectrolytes on enhanced chemiluminescence reaction in solution." Database accession no. PREV199698627596 XP002193761	1
Y	abstract & BIOTECHNOLOGY AND APPLIED BIOCHEMISTRY, vol. 22, no. 3, 1995, pages 249-260, ISSN: 0885-4513 ---	6, 9, 14-16
Y	US 5 747 254 A (PONTIUS BRIAN WYLIE) 5 May 1998 (1998-05-05) column 3, line 16 - line 29 column 9, line 45 - line 60 column 11, line 47 ---	6, 9-11, 14-16, 21, 22, 24, 25
Y	US 5 770 459 A (LELAND JONATHAN K ET AL) 23 June 1998 (1998-06-23) examples 10, 11 ---	6, 9-11, 14, 21, 22
A	WO 92 21975 A (ABBOTT LAB) 10 December 1992 (1992-12-10) example 15 -----	6, 9, 14

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 01/29390

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
US 5370993	A	06-12-1994	US 5136095 A	04-08-1992
			US 4812401 A	14-03-1989
			US 5405743 A	11-04-1995
			CA 1322067 A1	07-09-1993
			DE 3816953 A1	08-12-1988
			FR 2615621 A1	25-11-1988
			GB 2206206 A , B	29-12-1988
			JP 2657066 B2	24-09-1997
			JP 63314466 A	22-12-1988
EP 0895085	A	03-02-1999	EP 0895085 A1	03-02-1999
			JP 11118804 A	30-04-1999
US 5478729	A	26-12-1995	AU 2584495 A	29-11-1995
			CA 2188752 A1	09-11-1995
			DE 69504191 D1	24-09-1998
			DE 69504191 T2	11-02-1999
			EP 0757794 A1	12-02-1997
			ES 2133772 T3	16-09-1999
			JP 9512634 T	16-12-1997
			WO 9530151 A1	09-11-1995
US 5747254	A	05-05-1998	US 5474911 A	12-12-1995
			US 5015569 A	14-05-1991
			AU 7048791 A	26-06-1991
			CA 2069948 A1	02-06-1991
			EP 0503000 A1	16-09-1992
			WO 9108480 A1	13-06-1991
US 5770459	A	23-06-1998	US 5746974 A	05-05-1998
			AT 184320 T	15-09-1999
			AU 676665 B2	20-03-1997
			AU 2347892 A	11-02-1993
			CA 2112675 A1	21-01-1993
			DE 69229950 D1	14-10-1999
			DE 69229950 T2	09-03-2000
			EP 0594766 A1	04-05-1994
			EP 0854194 A2	22-07-1998
			ES 2137191 T3	16-12-1999
			GR 3031217 T3	31-12-1999
			JP 3053112 B2	19-06-2000
			JP 7508340 T	14-09-1995
			KR 212178 B1	02-08-1999
			WO 9301308 A1	21-01-1993
			AT 173542 T	15-12-1998
			AU 668085 B2	26-04-1996
			AU 1420692 A	07-09-1992
			AU 1530492 A	07-09-1992
			CA 2103674 A1	07-08-1992
			CN 1065339 A	14-10-1992
			CN 1064945 A	30-09-1992
			DE 69227624 D1	24-12-1998
			DE 69227624 T2	08-07-1999
			DK 570518 T3	02-08-1999
			EP 0570518 A1	24-11-1993
			EP 0877252 A2	11-11-1998
			ES 2126591 T3	01-04-1999
			GR 3029446 T3	28-05-1999

INTERNATIONAL SEARCH REPORT

Information on patent family members

Inter. Application No

PCT/US 01/29390

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
US 5770459	A		HK 1017970 A1	11-08-2000
			IE 920380 A1	12-08-1992
			IE 920381 A1	12-08-1992
			IL 100866 A	31-10-1995
			IL 100867 A	08-12-1995
			JP 3182515 B2	03-07-2001
			JP 11148901 A	02-06-1999
			JP 3128541 B2	29-01-2001
			JP 11125601 A	11-05-1999
			JP 3061416 B2	10-07-2000
			JP 6509412 T	20-10-1994
			JP 3013937 B2	28-02-2000
			JP 6508203 T	14-09-1994
			KR 226231 B1	15-10-1999
			NZ 241537 A	25-06-1993
			NZ 241538 A	25-02-1993
			WO 9214138 A1	20-08-1992
			WO 9214139 A1	20-08-1992
			US 5962218 A	05-10-1999
			US 5635347 A	03-06-1997
			US 6325973 B1	04-12-2001
<hr/>				
WO 9221975	A	10-12-1992	AT 161104 T	15-12-1997
			CA 2110296 A1	10-12-1992
			DE 69223510 D1	22-01-1998
			DE 69223510 T2	18-06-1998
			EP 0586574 A1	16-03-1994
			ES 2112320 T3	01-04-1998
			JP 6508214 T	14-09-1994
			US 5459078 A	17-10-1995
			WO 9221975 A1	10-12-1992
<hr/>				

THIS PAGE BLANK (USPTO)

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ **BLACK BORDERS**
- ☐ **IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- ☐ **FADED TEXT OR DRAWING**
- ☐ **BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- ☐ **SKEWED/SLANTED IMAGES**
- ☐ **COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- ☐ **GRAY SCALE DOCUMENTS**
- ☐ **LINES OR MARKS ON ORIGINAL DOCUMENT**
- ☐ **REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- ☐ **OTHER:** _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.

THIS PAGE BLANK (USPTO)